

## Article

# Determination of Florfenicol, Thiamfenicol and Chloramfenicol at Trace Levels in Animal Feed by HPLC–MS/MS

Rosa Elvira Gavilán <sup>1</sup>, Carolina Nebot <sup>1,\*</sup> , Ewelina Patyra <sup>2</sup>, Beatriz Vazquez <sup>1</sup> ,  
Jose Manuel Miranda <sup>1</sup>  and Alberto Cepeda <sup>1</sup>

<sup>1</sup> Department of Analytical Chemistry, Nutrition and Bromatology, Faculty of Veterinary Medicine, University of Santiago de Compostela, 27002 Lugo, Spain; rosaelviraclg@yahoo.es (R.E.G.); beatriz.vazquez@usc.es (B.V.); josemanuel.miranda@usc.es (J.M.M.); alberto.cepeda@usc.es (A.C.)

<sup>2</sup> Department of Hygiene of Animal Feedingstuffs, National Veterinary Research Institute, 24–100 Pulawy, Poland; ewelinapatyra@gmail.com

\* Correspondence: carolina.nebot@usc.es

Received: 20 March 2019; Accepted: 30 April 2019; Published: 7 May 2019



**Abstract:** Administration of florfenicol and thiamfenicol through medicated feed is permitted within the European Union, always following veterinary prescription and respecting the withdrawal periods. However, the presence of low levels of florfenicol, thiamfenicol, and chloramfenicol in non-target feed is prohibited. Since cross-contamination can occur during the production of medicated feed and according to Annex II of the European Regulation 2019/4/EC, the control of residue levels of florfenicol and thiamfenicol in non-target feed should be monitored and avoided. Based on all the above, a sensitive and reliable method using liquid chromatography tandem mass spectrometry was developed for the simultaneous detection of chloramfenicol, florfenicol, and thiamfenicol at trace levels in animal feed. Analytes were extracted from minced feed with ethyl acetate. Then, the ethyl acetate was evaporated, the residue was resuspended in Milli-Q water and the extract filtered. The method was in-house validated at carryover levels, with concentration ranging from 100 to 1000 µg/kg. The validation was conducted following the European Commission Decision 2002/657/EC and all performance characteristics were successfully satisfied. The capability of the method to detect amfenicols at lower levels than any prior perspective regulation literature guarantees its applicability in official control activities. The developed method has been applied to non-compliant feed samples with satisfactory results.

**Keywords:** non-target feed; florfenicol; thiamfenicol; chloramfenicol; HPLC–MS/MS; validation; swine

## 1. Introduction

Globalization permits food produced in one country to be sold in other countries, sometimes on an intercontinental level. However, globalization also contributes to competition between production companies. The final goal of any type of business is to have low production cost and high benefits. Low production cost is very important in food of animal origin, for which farmers fight with animal disease. The use of certain veterinary medicine is permitted to control, prevent, and treat illness. The most employed medicines, in this case, include antibiotics and antiparasitic agents. Antimicrobial medicines are sold as premixes, oral powders, oral solutions, injections, intramammary preparations pastes, oral pastes, boluses and intrauterine preparations [1]. Fenicols (chloramfenicol, thiamfenicol, and florfenicol) belong to this group of antibiotics; however, even if chloramfenicol is very effective on a broad spectrum, it is prohibited in food-producing animals within the European Union [2]. On the other hand, thiamfenicol and florfenicol can be administrated through feed,

but always respecting the withdrawal periods. Regulation 2019/4/EC [3] includes these two antibiotics as an active substance for medicated feed and also in Annex II of the Regulation, indicating that cross-contamination level should be investigated during the production of medicated feed.

Techniques such as phase sorptive extraction [4], indirect competitive enzyme-linked immunosorbent assays [5], molecularly imprinted solid-phase extraction [6], high performance liquid chromatography (HPLC) with ultraviolet (UV) detection [7], capillary electrophoresis [8], and QuEChERS [9] were employed for amfenicol analysis.

The European Decision Commission 2002/657/EC states that positive samples need to be confirmed with confirmatory methods, and that HPLC–MS/MS is a good technique for confirmatory methods [10]. The use of HPLC–MS/MS has been reported for amfenicol analysis in food matrices. However, when compared with other antibiotics, such as tetracyclines, sulfonamides or penicillin, there are few methods available for amfenicols. According to the ESVAC 2015 report, these three groups of antibiotics (tetracyclines, sulfonamides, and penicillin) accounted for approximately 70% of the total sales of antibiotics in the European Union. The reported HPLC–MS/MS methods include one described by Van de Riet et al. (2013) [11] for chloramfenicol, thiamfenicol, and florfenicol in fish muscle, one developed by Barreto et al. (2016) [12] to detect the same amfenicols in poultry, swine, bovine, and fish muscle, another published by Anderson et al. (2016) [13] for florfenicol and thiamfenicol in white-tailed deer, one described for detection in milk and honey [9], and the most recent one reported for detection in egg [14].

Regarding feed samples, Pietroni et al. (2014) [15] reported an HPLC–UV method for the quantification of florfenicol and thiamfenicol in medicated feed. Later, in 2017, a similar method was reported, but the feed extract was purified by thin layer chromatography (TLC) (Yang et al., 2017) [16]. For residue levels of amfenicols in feed, the technique most frequently employed is HPLC combined with different types of mass spectrometry. Between 30 and 300 active compounds (veterinary drugs, pesticides, and others) can be identified with low limits of detection (20 µg/kg). However, some of these methods are for screening [17–21] while others purify the extract with primary secondary amine (PSA) [22] or solid-phase extraction (SPE) [23].

To the best of the authors' knowledge, no method has been found in the scientific literature that is capable of analyzing all three amfenicols (chloramfenicol, thiamfenicol, and florfenicol) at the residual levels that fulfill the requirements of the European Commission Decision 2002/657/EC. Therefore, the aim of this research article is to report a simple extraction protocol followed by HPLC–MS/MS detection of amfenicols in feed samples from different animal species.

## 2. Results

The objective of this article is to describe a simple extraction protocol and an HPLC–MS/MS method to confirm the presence of amfenicols in feed samples at trace levels.

Figures 1 and 2 show multiple reaction monitoring (MRM) transitions of the different analytes in blank feed samples and in the same blank feed samples spiked with amfenicols to a final concentration of 100 µg/kg.

### 2.1. HPLC–MS/MS Conditions

Standard solutions of 1 µg/mL of florfenicol, thiamfenicol, chloramfenicol, and chloramfenicol-d<sub>5</sub> were employed to optimize their detection by the MS. The objective was to achieve a high signal intensity of representative ions through modifications in parameters such as curtain gas, ion spray voltage, source temperature, and curtain gas flow. Once precursor ions were selected for each amfenicol and optimized by manual tuning, automatic optimization was conducted (Table 1). This process permitted the evaluation of parameters such as entrance potential, collision cell potential, collision energy, and collision cell exit potential for four representative production ions. From these four ions, two were selected to conduct MRM analysis. Amfenicols were ionized by employing negative electrospray, due to the chemistry of the molecule; this ionization mode has been previously employed by other researchers in this field [14,17–24]. Acetonitrile was the first tested organic solvent as it is

preferential to methanol. Acetonitrile was combined with water and buffers (ammonium acetate, ammonium formate, and ammonium hydroxide), resulting in a good intensity signal. The best resolution between peaks and signal intensity was obtained with the combination of acetonitrile and water with ammonium formate and 0.1% of formic acid.

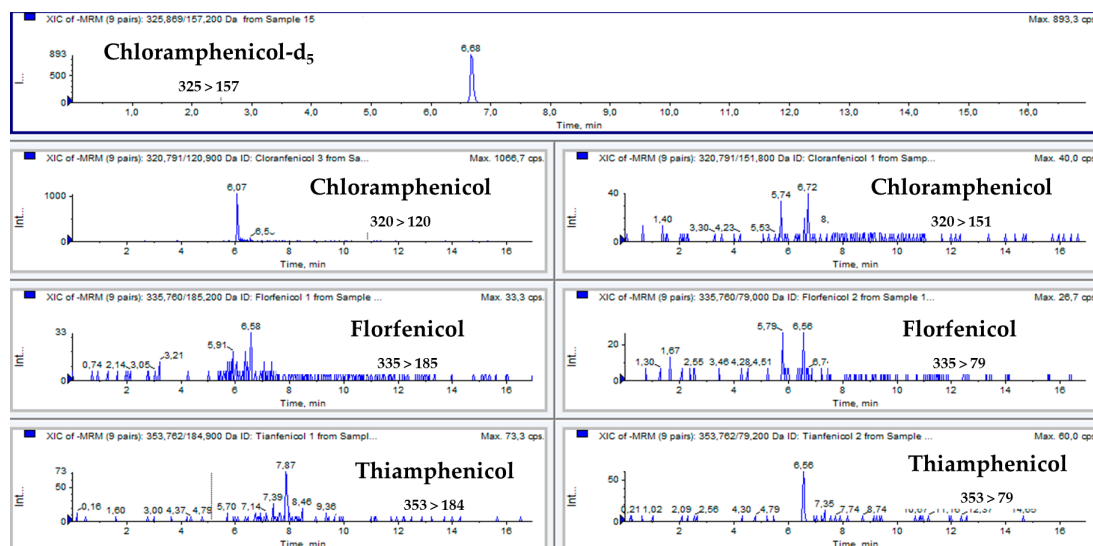


Figure 1. The total ion chromatogram (TIC) of a blank feed sample.

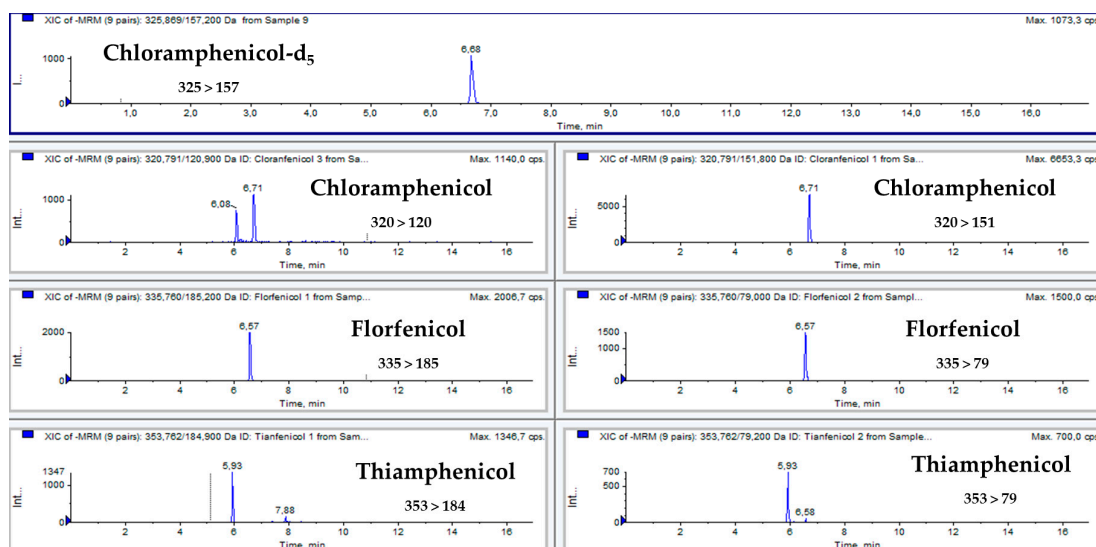


Figure 2. MRM chromatograms of feed samples spiked with amfenicols at the validation level of 200 µg/kg.

Table 1. Retention time (Rt), cone voltage (CV), collision energy, and precursor and product ions employed for ion identification.

Tetracycline	Rt (min)	m/z Transition	CV	Collision Energy
Chloramfenicol	6.71	320 > 151	46	26
Chloramfenicol	6.71	320 > 120	46	32
Thiamfenicol	5.93	353 > 184	56	20
Thiamfenicol	5.93	353 > 79	56	27
Florfenicol	6.57	335 > 185	61	23
Florfenicol	6.57	335 > 79	61	23
Chloramfenicol-d <sub>5</sub>	6.69	325 > 157	41	32

Following the requirement of the Decision 2002/657/EC, four identification points were achieved with the use of two MRM transitions for each amfenicol and the internal standard (IS). While both MRM transitions gave identification information, quantification was implemented from the MRM that gave the higher signal to noise ratio (S/N). The ions selected for chloramfenicol and florfenicol were also employed by Cronly et al. (2010) [25] and by Shinoda et al. (2011) [26] for detection in feed matrices. Similar results occurred between the selected ion for florfenicol and the work described by Shinoda et al. (2011) and Cronly et al. (2010), though the latter reference does not include florfenicol in its method. The same ions have also employed in other type of matrices, such as wastewater [27], manure [27], poultry tissues [28], pork, porcine liver, porcine kidney, beef, bovine liver, fish, chicken [29], milk, and honey [9]. The chromatography separation was performed with various C18 columns, including LiChrospher, Symmetry Shield RP18, ZORBAX Eclipse Plus, XTerra C18, and Hypersil C18-BD. For the present work, Sunfire C18 columns were selected.

## 2.2. Extraction

Independently of the matrix, most methods described in the scientific literature employ ethyl acetate to extract amfenicols [12,24,28,30]. Xiao et al. (2015) also employed ethyl acetate for poultry tissue but with a pressurized liquid extraction system [28]. Acetone and dichloromethane were selected by Van de Riet et al. (2013) to extract the drugs from fish muscle [11]. Acetonitrile with formic acid was employed by Boix et al. (2014) in feed samples and water combined with buffered acetonitrile by Leon et al. (2016) also for feed samples [21]. More sophisticated extraction techniques, such as QuEChERS, were employed by Liu et al. (2016) for milk and honey [9]. An immunoaffinity column was used by Luo et al. (2010) [31] for swine muscle and imprinted polymer by Ge et al. (2010) for animal tissue [6]. For the extraction of trace-level amfenicols from feed samples, 2 mL of water and 5 mL of ethyl acetate were employed with satisfactory results. However, after shaking for 20 minutes and centrifugation for the separation of the different phases, only 2 mL of the organic layer was evaporated. The evaporation of greater volumes (3, 3.5, and 4 mL) was investigated to increase the limit of detection (LOD), however, a higher matrix effect was observed, and the idea discarded.

## 2.3. Method Validation

The research group has developed, validated, and published various methods for the analysis of trace levels of veterinary drugs (coccidiostats, sulfonamides, and other antibiotics) in animal food and feed samples [32–35]. Validation for this type of sample and the matrix has typically been conducted following the European guideline (Decision 2002/657/EC) [10], which permits method implementation and comparisons in EU reference laboratories.

The Decision 2002/657/EC indicates that when no certificate reference material is available, trueness of measurements is assessed through recovery of additions of known amount of the analytes to a blank matrix. For amfenicols, recoveries were between −20% and +10% for the three investigated analytes (Table 2).

As with recovery, the precision under conditions of repeatability and reproducibility were evaluated using a standard addition method. Precision values were within the range provided by the European Decision; they ranged from 12% to 21% for chloramfenicol, 6.5% to 22% for thiamfenicol, and 12% to 19% for florfenicol. Currently, the presence of residue of amfenicols in non-target feed is prohibited; therefore, CC $\alpha$  and CC $\beta$  were calculated through employing formula for non-permitted substances. After peak identification, the area was plotted against the added concentration. CC $\alpha$  was the corresponding concentration at the y-intercept plus 2.33 times the standard deviation of the within-laboratory reproducibility. Similarly, for CC $\beta$  calculation, the peak area was plotted against the added concentration, and the CC $\alpha$  plus 1.64 times the standard deviation of the within-laboratory reproducibility of the mean measured content at the CC $\alpha$  equals the CC $\beta$  (2002/657/EC). While CC $\alpha$  values were 108, 140, and 110  $\mu\text{g/kg}$  for chloramfenicol, thiamfenicol, and florfenicol, respectively, CC $\beta$  values were 116, 180, and 122  $\mu\text{g/kg}$  for chloramfenicol, thiamfenicol, and florfenicol, respectively.

For the selected analytes, the S/N at the CC $\alpha$  was around 100; values for LOD and limit of quantification (LOQ) were calculated with the feed samples spiked with amfenicols. Results indicated that the LOD and LOQ values could reach 50  $\mu\text{g/kg}$ . However, to further validate the method, validation was conducted at 200  $\mu\text{g/kg}$  to confirm satisfactory S/N at the CC $\alpha$  level of each compound and to fulfill the EU requirements.

The technique itself is very selective/specific as it uses an MRM detection mode; the use of two MRM transitions reduces the detection of other interfering peaks. Selective/specific proxies were investigated with 20 blank feed samples for different animal species. The same 20 samples were spiked at 100  $\mu\text{g/kg}$  with amfenicols and other antibiotics (sulfadiazine, trimethoprim, tetracycline, and ciprofloxacin). Both the absence of interfering peaks at the Rt of the amfenicols in the two MRM transitions of each analyte and the correct identification of the analytes demonstrate the selectivity/specificity of the developed method.

**Table 2.** Recovery (%), repeatability (CV%), reproducibility (CV%), CC $\alpha$  ( $\mu\text{g/kg}$ ), CC $\beta$  ( $\mu\text{g/kg}$ ), LOD ( $\mu\text{g/kg}$ ) and LOQ ( $\mu\text{g/kg}$ ) of Chloramfenicol, Tiamfenicol and Florfenicol.

	Fortification Level ( $\mu\text{g/kg}$ )	Recovery ( $n = 18$ )	Repeatability ( $n = 6$ )	Reproducibility ( $n = 18$ )	CC $\alpha$	CC $\beta$	LOD	LOQ
Chloramfenicol	100	81	21	16	108	116	25	40
	200	88	12	16				
	300	89	15	19				
	Mean	86	16	17				
Tiamfenicol	100	81.41	6.5	22.35	140	180	75	100
	200	97.69	12.58	21.61				
	300	103.83	18.97	21.98				
	Mean	94.31	12.69	21.98				
Florfenicol	100	96.61	14.64	18.77	110	122	50	75
	200	94.680	12.97	12.32				
	300	90.342	13.22	15.22				
	Mean	93.88	13.61	15.44				

#### 2.4. Real Sample Collection and Analysis

Interlaboratory studies gave satisfactory results indicating the reliability and applicability of the developed method. Furthermore, the presence of the three amfenicols was investigated in 30 feed samples from feed mills. Florfenicol was the only amfenicol detected, and it was detected in one individual sample at a concentration of 0.36 mg/kg. This sample belonged to a group of four samples used to monitor florfenicol carryover after the production of a bath of medicated feed with 80 mg/kg of florfenicol. These results indicated that florfenicol carryover may occur during the manufacture of medicated feed and should be investigated in more detail. None of the samples collected from pig farms gave positive results.

### 3. Materials and Methods

#### 3.1. Chemicals, Reagents, and Stock Solutions

Florfenicol, thiamfenicol, chloramfenicol, and chloramfenicol-d<sub>5</sub> (purity > 98%) were purchased from Sigma-Aldrich (St. Louis, MO, USA). Chloramfenicol-d<sub>5</sub> was employed as internal standard (IS). Ethyl acetate and acetonitrile was obtained from Scharlau Chemie (Barcelona, Spain), and formic acid and ammonium acetate (purity >99% for analysis) were obtained from Acros Organics (Geel, Belgium). Purified water was prepared in-house with a Milli-Q water system from Millipore (Bedford, MA, USA).

Nitrogen gas was generated using an in-house nitrogen generator from Peak Scientific Instruments, Ltd (Chicago, IL, USA).

In 20 mL volumetric flasks,  $20 \pm 0.01$  mg of each analyte (florfenicol, thiamfenicol, chloramfenicol, and chloramfenicol- $d_5$ ) were precisely weighed to prepare stock solutions of individual compounds in acetonitrile (to yield a final concentration of 1 mg/mL; the purity was considered when calculating specific concentrations). To prepare the intermediate stock solution (10  $\mu$ g/mL), the individual solutions were mixed and diluted with acetonitrile. Each day, a working stock solution (1  $\mu$ g/mL) was freshly prepared by diluting the intermediate stock solution with acetonitrile. All standards solutions were stored in the dark at  $-18^\circ\text{C}$  for no longer than three months.

### 3.2. Conditions for HPLC–MS/MS Analysis

The HPLC–MS/MS analyses were performed using a 1100 HPLC from Agilent Technologies (Waldbronn, Germany) attached to a QTRAP 2000™ MS from Applied Biosystems/MDS-Sciex (Toronto, Canada). The software Analyst 1.4.1 from Applied Biosystems (Toronto, Canada) was employed to control the system.

The analysis of the extracts with amfenicols was achieved with a Sunfire C18 (3.5  $\mu$ m 2.1  $\times$  150 mm) HPLC column from Waters (Milford, PA, USA), water with formic acid (185  $\mu$ L), and 370  $\mu$ L of ammonium formate (6.3% in water; mobile phase A), and acetonitrile (mobile phase B). Mobile phase components A and B were freshly prepared with each batch of sample. The solvents were mixed at a constant flow rate of 250  $\mu$ L/min, the column temperature was maintained at  $50^\circ\text{C}$ , and the injection volume was 40  $\mu$ L. The gradient program was as follows: 0 min, 90% A; 1 min, 50% A; 5 min, 20% A; 7 min, 90% A; 17 min, 90% A.

The MS analysis was performed using negative electrospray ionization mode (ESI). The dwell time was 150 ms between transitions. The system uses nitrogen as nebulized gas set up at 50 psi for ion source gases 1 and 2. The source temperature was  $475^\circ\text{C}$ , the ion spray voltage was  $-4500$  V, and the curtain gas flow was 25 psi. Analytes were identified by retention time (Rt) and two multiple reaction monitoring (MRM) transitions. Table 1. compiles the MS parameters employed for the identification of each analyte.

### 3.3. Sample Extraction

To confirm the absence of chloramfenicol, feed samples were sent to an accredited laboratory. Once the results were obtained, these samples were employed as blank feed samples to prepare matrix-matched calibration.

Quantification of the analytes was performed with matrix-matched samples spiked with the analytes at concentrations of 100, 200, 300, 400, and 1000  $\mu$ g/kg. Samples were ground on a Minimoka GR-020 grinder (Lleida, Spain), and 1 g was transferred to a 15 mL Falcon conical-bottom tube. Then, 2 mL of Milli-Q water and 5 mL of ethyl acetate were added to the tube. The mixture was mixed with an IKA Minishaker MS2 (Staufen, Germany) for 20 s, shaken for 30 min at 200 rpm on a New Brunswick Scientific G25 orbital shaker (NJ, USA), and centrifuged at 7000 rpm for 10 minutes on a Kokusan H-103N centrifuge (Tokyo, Japan). Then, a volume of 2 mL was transferred to a conical glass tube and evaporated to dryness with a stream of nitrogen on a TurboVap® II evaporator from Zymark (MA, USA). The dry residue was dissolved in 500  $\mu$ L of Milli-Q water, filtered with a GHP Acrodisc syringe filter (0.2  $\mu$ m; Waters Corporation, MA, USA), and transferred to an HPLC amber vial with an insert. The extract was stored at  $-20^\circ\text{C}$  until analysis, which was conducted within a day. Figure 1 shows the total ion chromatogram (TIC) of a blank feed sample, and Figure 2 shows MRM chromatograms of feed samples spiked with amfenicols at the validation level of 200  $\mu$ g/kg.

### 3.4. Method Validation

Linearity range, recovery, precision (repeatability and reproducibility), selectivity/specificity, limit of detection (LOD), limit of quantification (LOQ), decision limit ( $CC\alpha$ ), and detection capability



(CC $\beta$ ) of the developed method were obtained, along with validation of the method. The validation was conducted following the criteria included in the Commission Decision 657/2002/EC. Blank feed samples were spiked with the amfenicols at 100, 200, 300, 400, and 1000  $\mu\text{g/kg}$  for linearity verification. Calibration curves were prepared over four different days; for each day, peak area was correlated against analyte concentrations for linear regression analysis. The accuracy of the method could not be determined as certified reference materials were not available. Therefore, trueness and precision were obtained with blank feed samples spiked at 100, 200, and 300  $\mu\text{g/kg}$ . Six replicates of each concentration were extracted and analyzed on the same day to obtain intra-day precision (repeatability). The same procedure was repeated over three different days for inter-day precision (reproducibility).

The selectivity/specificity was evaluated with blank feed samples spiked and non-spiked with amfenicols at the validation level (100  $\mu\text{g/kg}$ ). The different feed samples were provided by the manufacturers. Selectivity/specificity was evaluated with feed samples for different animal species (swine, poultry, and cattle).

### 3.5. Sample Collection and Analysis

Feed producers provided non-targeted feed samples ( $n = 30$ ) to the laboratory. Four of these samples belonged to a feed mill and were collected after being manufactured in the same production line of medicated feed and after the cleaning batches to evaluate carryover contamination. The other feed samples were collected in pig farms and were feed employed for pig crowing. Once in the laboratory, all samples were ground and kept in plastic bottles in the dark until analysis. Interlaboratory samples from 2018 RIKILT proficiency tests were also conducted.

## 4. Conclusions

The article describes a simple and rapid confirmatory method based on HPLC–MS/MS for the simultaneous identification and quantification of residue of chloramfenicol, thiamfenicol and florfenicol in non-target feed. Since the method was validated following the EU guidelines and fulfilled the requirements of the Decision, the method can be applied by different laboratories, including reference laboratories.

**Author Contributions:** Investigation, R.E.G. and C.N.; Methodology, R.E.G. and E.P.; Supervision, J.M.M. and A.C.; Validation, C.N.; Writing—original draft, C.N. and B.V.

**Funding:** This research was funded by FEADER (The European Agricultural Fund for Rural Development (EAFRD)), grant number 2018/001B and ED431C 2018/05.

**Conflicts of Interest:** The authors do not have any conflict of interest.

## References

1. ESVAC (2015) Fifth ESVAC Report—Sales of Veterinary Antimicrobial Agents in 26 EU/EEA Countries in 2013. Available online: [www.ema.europa.eu/docs/en\\_GB/document\\_library/Report/2015/10/WC500195687.pdf](http://www.ema.europa.eu/docs/en_GB/document_library/Report/2015/10/WC500195687.pdf) (accessed on 29 January 2017).
2. Europe Commission. Regulation 37/2010. Commission Regulation (EU) No 37/2010 of 22 December 2009 on pharmacologically active substances and their classification regarding maximum residue limits in foodstuffs of animal origin. *Off. J. Eur. Union* **2010**, *15*, 1–72.
3. Europe Commission. Regulation 2019/4. Commission Regulation (EU) 2019/4 of the European Parliament and of the Council of 11 December 2018 on the manufacture, placing on the market and use of medicated feed, amending Regulation (EC) No 1831/2003 of the European Parliament and of the Council and repealing Council Directive 90/167/EEC (Text with EEA relevance). *Off. J. Eur. Union* **2019**, *4*, 1–23.
4. Samanidou, V.; Kaltzi, I.; Kabir, A.; Furton, K.G. Simplifying sample preparation using fabric phase sorptive extraction technique for the determination of benzodiazepines in blood serum by high performance liquid chromatography. *Biomed. Chromatogr.* **2016**, *30*, 829–836. [[CrossRef](#)] [[PubMed](#)]

5. An, L.; Wang, Y.; Pan, Y.; Tao, Y.; Chen, D.; Liu, Z.; Yuan, Z. Development and validation of a sensitive indirect competitive enzyme-linked immunosorbent assay for the screening of florfenicol and Tiamfenicol in edible animal tissue and feed. *Food Anal. Method* **2016**, *9*, 2434–2443. [\[CrossRef\]](#)
6. Ge, S.; Yan, M.; Cheng, X.; Zhang, C.; Yu, J.; Zhao, P.; Gao, W. On-line molecular imprinted solid-phase extraction flow-injection fluorescence sensor for determination of florfenicol in animal tissues. *J. Pharm. Biomed.* **2010**, *52*, 615–619. [\[CrossRef\]](#) [\[PubMed\]](#)
7. Hayes, J. Determination of florfenicol in fish feeds at high inclusion rates by HPLC-UV. *J. AOAC Int.* **2013**, *96*, 7–11. [\[CrossRef\]](#) [\[PubMed\]](#)
8. Kowalski, P.; Konieczna, L.; Chmielewska, A.; Olędzka, I.; Plenis, A.; Bieniecki, M.; Lamparczyk, H. Comparative evaluation between capillary electrophoresis and high-performance liquid chromatography for the analysis of florfenicol in plasma. *J. Pharm. Biomed.* **2005**, *39*, 983–989. [\[CrossRef\]](#)
9. Liu, H.Y.; Lin, S.L.; Fuh, M.R. Determination of chloramfenicol, Tiamfenicol and florfenicol in milk and honey using modified QuEChERS extraction coupled with polymeric monolith-based capillary liquid chromatography tandem mass spectrometry. *Talanta* **2016**, *150*, 233–239. [\[CrossRef\]](#) [\[PubMed\]](#)
10. Europe Commission. Commission Decision 2002/657/EC. Commission Decision of 12 August 2002 implementing Council Directive 96/23/EC concerning the performance of analytical methods and the interpretation of results. *Off. J. Eur. Union* **2002**, *221*, 8–36.
11. Van de Riet, J.M.; Potter, R.A.; Christie-Fougere, M.; Burns, B.G. Simultaneous determination of residues of chloramfenicol, tiamfenicol, florfenicol, and florfenicol amine in farmed aquatic species by liquid chromatography/mass spectrometry. *J. AOAC Int.* **2003**, *86*, 510–514. [\[PubMed\]](#)
12. Barreto, F.; Ribeiro, C.; Hoff, R.B.; Dalla Costa, T. Determination of chloramfenicol, Tiamfenicol, florfenicol and florfenicol amine in poultry, swine, bovine and fish by liquid chromatography-tandem mass spectrometry. *J. Chromatogr. A* **2016**, *1449*, 48–53. [\[CrossRef\]](#)
13. Anderson, S.C.; Subbiah, S.; Gentles, A.; Austin, G.; Stonum, P.; Brooks, T.A.; Smith, E.E. Qualitative and quantitative drug residue analyses: Florfenicol in white-tailed deer (*Odocoileus virginianus*) and supermarket meat by liquid chromatography tandem-mass spectrometry. *J. Chromatogr. B* **2016**, *1033*, 73–79. [\[CrossRef\]](#) [\[PubMed\]](#)
14. Xie, X.; Wang, B.; Pang, M.; Zhao, X.; Xie, K.; Zhang, Y.; Dai, G. Quantitative analysis of chloramfenicol, tiamfenicol, florfenicol and florfenicol amine in eggs via liquid chromatography-electrospray ionization tandem mass spectrometry. *Food Chem.* **2018**, *269*, 542–548. [\[CrossRef\]](#)
15. Pietro, W.J.; Woźniak, A.; Pasik, K.; Cybulski, W.; Krasucka, D. Amfenicols stability in medicated feed—development and validation of liquid chromatography method. *Bull. Vet. Inst. Pulawy* **2014**, *58*, 621–629. [\[CrossRef\]](#)
16. Yang, J.; Sun, G.; Qian, M.; Huang, L.; Ke, X.; Yang, B. Development of a high-performance liquid chromatography method for the determination of florfenicol in animal feedstuffs. *J. Chromatogr. B* **2017**, *1068*, 9–14. [\[CrossRef\]](#) [\[PubMed\]](#)
17. Boix, C.; Ibáñez, M.; Sancho, J.V.; León, N.; Yúsà, V.; Hernández, F. Qualitative screening of 116 veterinary drugs in feed by liquid chromatography–high resolution mass spectrometry: Potential application to quantitative analysis. *Food Chem.* **2014**, *160*, 313–320. [\[CrossRef\]](#)
18. Robert, C.; Gillard, N.; Brasseur, P.Y.; Ralet, N.; Dubois, M.; Delahaut, P. Rapid multiresidue and multi-class screening for antibiotics and benzimidazoles in feed by ultra-high performance liquid chromatography coupled to tandem mass spectrometry. *Food Control* **2015**, *50*, 509–515. [\[CrossRef\]](#)
19. Aguilera-Luiz, M.M.; Romero-González, R.; Plaza-Bolaños, P.; Vidal, J.L.M.; Frenich, A.G. Wide-scope analysis of veterinary drug and pesticide residues in animal feed by liquid chromatography coupled to quadrupole-time-of-flight mass spectrometry. *Anal. Bioanal. Chem.* **2013**, *405*, 6543–6553. [\[CrossRef\]](#) [\[PubMed\]](#)
20. Gómez-Pérez, M.L.; Romero-González, R.; Martínez Vidal, J.L.; Garrido Frenich, A. Analysis of veterinary drug and pesticide residues in animal feed by high-resolution mass spectrometry: Comparison between time-of-flight and Orbitrap. *Food Addit. Contam. A* **2015**, *32*, 1637–1646. [\[CrossRef\]](#)
21. León, N.; Pastor, A.; Yúsà, V. Target analysis and retrospective screening of veterinary drugs, ergot alkaloids, plant toxins and other undesirable substances in feed using liquid chromatography–high resolution mass spectrometry. *Talanta* **2016**, *149*, 43–52. [\[CrossRef\]](#)



22. Boscher, A.; Guignard, C.; Pellet, T.; Hoffmann, L.; Bohn, T. Development of a multi-class method for the quantification of veterinary drug residues in feedingstuffs by liquid chromatography-tandem mass spectrometry. *J. Chromatogr. A* **2010**, *1217*, 6394–6404. [[CrossRef](#)]
23. Piatkowska, M.; Jedziniak, P.; Zmudzki, J. Multiresidue method for the simultaneous determination of veterinary medicinal products, feed additives and illegal dyes in eggs using liquid chromatography–tandem mass spectrometry. *Food Chem.* **2016**, *197*, 571–580. [[CrossRef](#)]
24. Zhang, S.; Liu, Z.; Guo, X.; Cheng, L.; Wang, Z.; Shen, J. Simultaneous determination and confirmation of chloramfenicol, tiamfenicol, florfenicol and florfenicol amine in chicken muscle by liquid chromatography–tandem mass spectrometry. *J. Chromatogr. B* **2008**, *875*, 399–404. [[CrossRef](#)]
25. Cronly, M.; Behan, P.; Foley, B.; Malone, E.; Martin, S.; Doyle, M.; Regan, L. Rapid multi-class multi-residue method for the confirmation of chloramfenicol and eleven nitroimidazoles in milk and honey by liquid chromatography-tandem mass spectrometry (LC-MS). *Food Addit. Contam.* **2010**, *27*, 1233–1246. [[CrossRef](#)] [[PubMed](#)]
26. Shinoda, N.; Kojima, F.; Sugiura, K. Simultaneous determination of residues of chloramfenicol and florfenicol in animal feed by liquid chromatography tandem mass spectrometry. *J. Resid. Sci. Technol.* **2011**, *8*, 125–129.
27. Wei, R.; Ge, F.; Chen, M.; Wang, R. Occurrence of ciprofloxacin, enrofloxacin, and florfenicol in animal wastewater and water resources. *J. Environ. Qual.* **2012**, *41*, 1481–1486. [[CrossRef](#)]
28. Xiao, Z.; Song, R.; Rao, Z.; Wei, S.; Jia, Z.; Suo, D.; Fan, X. Development of a subcritical water extraction approach for trace analysis of chloramfenicol, tiamfenicol, florfenicol, and florfenicol amine in poultry tissues. *J. Chromatogr. A* **2015**, *1418*, 29–35. [[CrossRef](#)]
29. Chou, K.Y.; Cheng, T.Y.; Chen, C.M.; Hung, P.L.; Tang, Y.Y.; Chung-Wang, Y.J.; Shih, Y.C. Simultaneous determination of residual tiamfenicol and florfenicol in foods of animal origin by HPLC/electrospray ionization-MS/MS. *J. AOAC Int.* **2009**, *92*, 1225–1232. [[PubMed](#)]
30. Peng, L.I.; Yueming, Q.I.U.; Huixia, C.A.I.; Ying, K.O.N.G.; Yingzhang, T.A.N.G.; Daning, W.A.N.G.; Mengxia, X.I.E. Simultaneous determination of chloramfenicol, tiamfenicol, and florfenicol residues in animal tissues by gas chromatography/mass spectrometry. *Chin. J. Chromatogr.* **2006**, *24*, 14–18.
31. Luo, P.; Chen, X.; Liang, C.; Kuang, H.; Lu, L.; Jiang, Z.; Shen, J. Simultaneous determination of thiamfenicol, florfenicol and florfenicol amine in swine muscle by liquid chromatography–tandem mass spectrometry with immunoaffinity chromatography clean-up. *J. Chromatogr. B* **2010**, *878*, 207–212. [[CrossRef](#)] [[PubMed](#)]
32. Nebot, C.; Iglesias, A.; Regal, P.; Miranda, J.; Cepeda, A.; Fente, C. Development of a multi-class method for the identification and quantification of residues of antibiotics, coccidiostats and corticosteroids in milk by liquid chromatography–tandem mass spectrometry. *Int. Dairy J.* **2012**, *22*, 78–85. [[CrossRef](#)]
33. Nebot, C.; Guarddon, M.; Seco, F.; Iglesias, A.; Miranda, J.M.; Franco, C.M.; Cepeda, A. Monitoring the presence of residues of tetracyclines in baby food samples by HPLC-MS/MS. *Food Control* **2014**, *46*, 495–501. [[CrossRef](#)]
34. Gavilán, R.; Nebot, C.; Miranda, J.; Martín-Gómez, Y.; Vázquez-Belda, B.; Franco, C.; Cepeda, A. Analysis of tetracyclines in medicated feed for food animal production by HPLC-MS/MS. *Antibiotics* **2016**, *5*, 1. [[CrossRef](#)] [[PubMed](#)]
35. Gavilán, R.E.; Nebot, C.; Patyra, E.; Miranda, J.M.; Franco, C.M.; Cepeda, A. Simultaneous analysis of coccidiostats and sulphonamides in non-target feed by HPLC-MS/MS and validation following the Commission Decision 2002/657/EC. *Food Addit. Contam. A* **2018**, *35*, 1093–1106. [[CrossRef](#)] [[PubMed](#)]

